

E-ISSN: 2229-7677 • Website: www.ijsat.org • Email: editor@ijsat.org

Environmental and Public Health Impacts of Heavy Metal Contamination in Coastal and Freshwater Edible Species

Ms. Jumoke Elizabeth Ajayi

Bemidji State University

Abstract

Heavy metal contamination in aquatic environments has become a critical global concern, posing significant risks to both ecosystem and human health. This study examines the environmental and public health impacts of heavy metal pollution in coastal and freshwater edible species. It explores the major sources of contamination including industrial discharge, agricultural runoff, mining activities, and atmospheric deposition and how these pollutants bioaccumulate through aquatic food webs. The research highlights key metals of concern such as mercury (Hg), cadmium (Cd), lead (Pb), arsenic (As), and chromium (Cr), which are frequently detected at concentrations exceeding international safety limits. Their persistence and toxicity contribute to biodiversity loss, ecosystem imbalance, and adverse human health outcomes including neurotoxicity, renal dysfunction, and carcinogenic effects. It also reviews existing regulatory frameworks and emphasizes the need for more stringent control measures, improved waste management, and public awareness campaigns. Overall, this paper explains the urgent necessity of integrating environmental management and food safety policies to safeguard both aquatic life and human populations dependent on these resources for nutrition and livelihood.

Keywords: Heavy metals, coastal ecosystems, freshwater species, bioaccumulation, food safety, public health, environmental pollution, mercury, cadmium, lead, risk assessment, aquatic contamination.

1.0 Introduction

Water is an essential requirement for all life forms on Earth. Clean water is vital for maintaining health, since contaminated water can jeopardize public health through direct or indirect exposure to hazardous chemicals [1]. Environmental contamination has been intensified by the industrial revolution and human activities [2]. Substantial pollutant emissions into the water have caused considerable threats to coastal ecosystems [3]. Heavy metals (HMs) are very detrimental environmental pollutants due to their chronic toxicity, non-biodegradability, and propensity for bioaccumulation [4]. Heavy metals can be transmitted and biomagnified across food systems, posing significant risks to human health [5, 6]. There is a significant demand for effective monitoring and surveillance of heavy metal concentrations in the marine environment [7]. Concerns over the concentration of heavy metals (HMs), along with their impacts, distribution, and environmental origins, are currently being raised at local, regional, and national levels [8].



E-ISSN: 2229-7677 • Website: www.ijsat.org • Email: editor@ijsat.org

The bioaccumulation patterns of heavy metals, including mercury (Hg), arsenic (As), nickel (Ni), cobalt (Co), copper (Cu), cadmium (Cd), and chromium (Cr), significantly impact the lifespan of most organisms [9]. Heavy metals from various distribution sources adversely affect marine biota [10]. These heavy metals affect beneficial creatures, including fish and various invertebrates [10]. Heavy metals from the ambient water and food sources accumulate in marine organisms [11]. In specific instances, elevated concentrations of heavy metals in marine ecosystems are directly associated with environmental pollution. Numerous research investigations indicate that the levels of heavy metal bioaccumulation varied significantly among marine species. The disparities in heavy metal accumulation among aquatic creatures may be attributed to their distinct habitats, dietary habits, and trophic levels [12, 13].

This review aims to elucidate the distribution of heavy metal sources in aquatic ecosystems and addresses heavy metal contamination in marine food constituents such as fish. Moreover, the impact of these components on the environment and human health is comprehensively examined to elucidate the physiological and molecular mechanisms associated with the utilization of metallic poisons in aquatic food sources. These management measures are closely associated with human population safety by preventing or reducing the passage of heavy metal contaminants from the aquatic environment to the food chain.

2.0 Sources and Pathways of Heavy Metal Contamination

Cadmium (Cd), zinc (Zn), mercury (Hg), arsenic (As), silver (Ag), chromium (Cr), copper (Cu), iron (Fe), and platinum (Pt) are all heavy metals (HMs) [14]. They are elements with a higher density and atomic mass that can harm people and the environment. One of the worst things that may happen to the ecosystem is heavy metal pollution in water, which affects plants, animals, and people [15, 16]. Heavy metals are dangerous even in small amounts because they don't break down in the environment [17, 18]. There are three classes of metals and metalloid ions. The first group has metals including mercury, cadmium, and lead that are dangerous even at low levels. The second group of metals is less harmful (bismuth, indium, arsenic, thallium), and the third group includes important metals like zinc, cobalt, copper, iron, and selenium, which are only harmful at high levels [19]. Heavy metals build up in the earth, human and animal tissues because they are absorbed or, in some cases, breathed in, or because of accidents or bad handling [20].

The primary contributors to increased environmental toxicity due to heavy metals are human and manmade influences. Natural sources of heavy metals (HMs) are things like soil that has blown in by the wind, forest fires, volcanic eruptions, biogenic processes, and marine salt. Mining, pesticides, fertilizers, and herbicides are all human-made sources of HMs contamination. Irrigating crop fields with industrial and sewage water is also a source of HMs contamination (Fig. 1). Trace amounts of heavy metals in fertilizers are a major source of heavy metal contamination in our food. Inappropriate industrial waste management, traffic pollution, use of lead (Pb) as fuel antiknock, aerosol cans, metallurgy and smelting, discharge of sewage and construction materials are the anthropogenic practices responsible for HMs contamination [17-20].



E-ISSN: 2229-7677 • Website: www.ijsat.org • Email: editor@ijsat.org

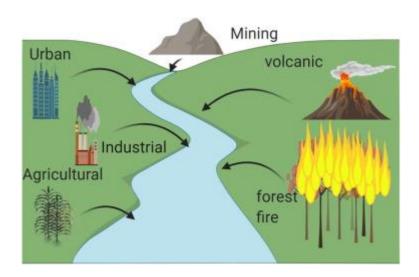


Figure 1: Source of HMs

The pharmaceutical production, paper and pulp preservatives, agricultural, chlorine and caustic soda sectors all let mercury (Hg) into the air [21]. Cadmium is present in soils and rocks, including coal and mineral fertilizer. Cadmium (Cd) is commonly utilized in electroplating for several purposes, including batteries, pigments, textiles, and metal coatings [22]. All of these actions are to blame for the higher levels of HMs pollution in the environment.

3.0 Heavy Metals of Concerns

Heavy metal pollution is becoming as a worldwide concern. Heavy metals can infiltrate fish via three pathways: the gills, the integumentary surface, and the gastrointestinal system [23]. Fish juveniles and larvae exhibit rapid growth, with their body length and mass directly correlated to appropriate temperature and adequate food availability [24]. Conversely, fish development is impeded by contaminated diet rich in heavy metals. A conspicuous indicator of metal toxicity in fish is growth suppression. Consequently, heavy metal concentrations in tissues induce a range of metabolic, physiological, and histological alterations in fish and other freshwater species by modifying different enzymes and metabolites [25].

The feeding process varies among fish species due to factors such as developmental influences, psychological issues, and lifetime. Heavy metals accumulate in the tissues of fish inhabiting contaminated environments [26, 27]. Metal intensity, expression duration, metal absorption, environmental variables (temperature, pH, hardness, and salinity), and intrinsic agents, such as fish age and feeding activities are all factors in the selection of body organs for HMs deposition. Metals predominantly concentrate in the kidneys, gills, and liver [28, 29]. Zinc accumulates in fish gills, obstructing the oxygen flow to tissues and resulting in hypoxia, ultimately causing mortality [30]. Nonetheless, a decrease in water pH may result in the mobilization and release of heavy metals into the water column, jeopardizing marine animals, including crustaceans and insects [31]. These poisonous sediments exterminate benthic species and diminish food availability for the larger organism.

At moderate levels, heavy metals present in the environment and food are essential for good health; nevertheless, excessive quantities can be detrimental or hazardous [32]. Their toxicity can diminish energy and impact the brain, lungs, kidneys, liver, blood, and other essential organs. Prolonged exposure ultimately leads to degenerative physical, tissue, and neurological processes that mimic diseases such as Alzheimer's, Parkinson's, muscular dystrophy, and multiple sclerosis [33, 34, 35]. Acute lead (Pb)



E-ISSN: 2229-7677 • Website: www.ijsat.org • Email: editor@ijsat.org

exposure can induce appetite loss, headaches, hypertension, stomach discomfort, renal dysfunction, fatigue, insomnia, arthritis, hallucinations, and vertigo. Mercury toxicity results in acrodynia or pink disease [36]. Increased mercury exposure may impact cerebral structure and induce shyness, tremors, cognitive decline, irritability, and visual or auditory impairments [37]. Inhalation of elevated levels of metallic mercury vapors for a brief duration may lead to pulmonary injury, emesis, diarrhea, nausea, dermal eruptions, and hypertension [38]. Organic mercury toxicity signs and symptoms include depression, memory problems, tremors, fatigue, headache, and hair loss. Because these signs and symptoms are frequently associated with other diseases, circumstances may be difficult to recognize [39, 40].

4.0 Contamination in Coastal and Freshwater Edible Species

The evaluation of bioaccumulation is a critical indicator for monitoring the geochemical cycle of heavy metals within aquatic ecosystems. The toxicological effects and oxidation states of heavy metals differ according to their specific forms and kinds. Chromium (Cr) typically occurs in six oxidative states (+1 to +6), with hexavalent Cr causing detrimental consequences in fish [41]. Fish in aquatic systems contaminated with heavy metals present a significant risk, as they accumulate metals in several critical bodily tissues (gills, liver, kidneys, skin, muscle, etc.), as depicted in Figure 2. Fish necessitate increased energy, derived from stored foods such as proteins, lipids, and carbs, to adapt to this stressful condition. Certain metals (As, Cd, Cr, Cu, Fe, Hg, Ni, Pb, Zn) possess redox potential and react to generate reactive oxygen species (ROS), which are crucial for sustaining specific physiological functions in fish. Reactive oxygen species (ROS) serve as a marker of oxidative stress, which impedes cellular function by destroying proteins, lipids, and DNA. Heavy metals bioaccumulate in various aquatic creatures via the food chain, leading to significant human health concerns upon the eating of contaminated fish [42, 43, 44, 45, 46].



E-ISSN: 2229-7677 • Website: www.ijsat.org • Email: editor@ijsat.org

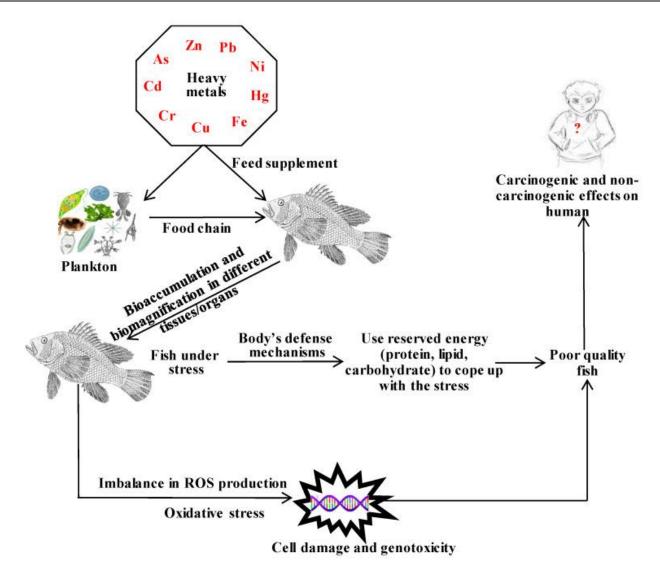


Figure 2: Mechanism of the contaminations of Heavy metals

Arsenic is one of the most hazardous heavy metals that contaminates aquatic ecosystems through numerous natural and anthropogenic activities [47]. Inorganic arsenic has been observed to be more hazardous than organic forms [48, 49]. Accumulation occurs in several organs of fish at disparate rates [50, 51]. The maximum concentration of As $(10.04 \pm 2.99 \ \mu g/g)$ was detected in the liver, while the minimum $(3.74 \pm 3.38 \ \mu g/g)$ was found in muscle after 20 days of exposure in Oreochromis niloticus [52]. Numerous studies have indicated that arsenic exposure adversely affects fish, resulting in diminished growth and productivity, alterations in hemato-biochemical parameters, hormonal dysregulation, histopathological abnormalities, delays in embryonic and larval development, and other disorders [53, 54, 55, 56, 57].

Furthermore, toxicity substantially impacted the hematological and immunological parameters of various fish species [58, 59, 60, 61]. An elevated dosage of arsenic led to increased mucus secretion, atypical swimming behavior, and impaired equilibrium in Anabas testudineus and Danio rerio [62, 63]. Induced various cytotoxic and genotoxic effects in medaka, Oryzias latipes [64]. Furthermore, pollution



E-ISSN: 2229-7677 • Website: www.ijsat.org • Email: editor@ijsat.org

impaired the reproductive functions of fish by hindering gametogenesis, thereby diminishing the quality and quantity of sperm and ova, as well as reducing fertilization and hatching success [65, 66, 67].

Cadmium is very poisonous and carcinogenic to humans and several animals, including fish. The Agency for Toxic Substances and Disease Registry of the United States identifies this metal as the eighth most dangerous substance. Numerous studies indicate that the aquatic environment is markedly polluted with cadmium (Cd) [68, 69, 70]. The assimilation and bioaccumulation of this hazardous metal have been seen in various aquatic organisms. Cadmium toxicity has caused dysfunction in various vital organs of fish, including the liver, kidneys, and gills, thereby impacting their physiology and inhibiting growth. Furthermore, Cd modifies hematological parameters by disrupting iron metabolism and inducing anemic conditions [71, 72]. Cadmium inhibits antioxidant enzymes, leading to lipid peroxidation in animals [73, 74]. Furthermore, cadmium exposure adversely impacts the reproductive efficacy of fish by reducing sperm lobule size, inducing testicular fibrosis, and diminishing sperm motility and viability [75, 76, 77].

Chromium is a pervasive metal that degrades environmental quality, originating from several industrial sources [78, 79]. Numerous investigations have documented the bioaccumulation of chromium in many organs of Cyprinus carpio [80, 81], and Carassius auratus [82], O. aureus [83]. Chromium poisoning disrupts the physiological activities of fish, leading to several inflammatory reactions and organ system failures [84, 85, 86, 87]. Furthermore, chromium toxicity markedly modifies the protein, lipid, and glycogen levels in the muscle, liver, and gills of Labeo rohita and C. mrigala, induces hepatic stress in C. auratus, disrupts the functions of vital organs (liver, kidney) in Ctenopharyngodon idella, and leads to dysfunctions of the endocrine system in various freshwater fish species. Chromium was observed to modify the blood profile, leading to cellular and nuclear anomalies in Pangasianodon hypophthalmus [87]. Numerous investigations indicated that elevated chromium levels in fish diets markedly reduced the development and feed efficiency of various fish species [88]. Furthermore, prolonged exposure to chromium led to reproductive complications in fish, including reduced spawning success [89, 90], and impaired oocyte development [91].

Copper is a substantial pollutant in aquatic systems that induces stress in aquatic species and markedly impedes the growth and physiology of fish [92, 93, 94]. Numerous studies indicate that the liver is the primary organ that accumulates a substantial amount of copper relative to other organs [95, 96, 97]. Excess copper in the fish diet diminished appetite, hence adversely impacting feed consumption and development in fish [98]. Furthermore, Cu poisoning led to malformations in reproductive organs and significantly decreased the GSI, fecundity, fertilization, and hatching rates of various fish species [99].

Manganese is frequently seen in diverse contexts. Manganese was discovered to leach into aquatic environments via multiple human activities [100]. Various factors, such as fish species, age, and water quality, can influence Mn toxicity in fish [101]. Mn toxicity diminishes when water hardness increases [100]. The bioaccumulation of manganese in the liver, gills, and muscles of Argyrosomus japonicus disrupted glucose metabolism and modified the ionic composition of blood plasma [102]. Manganese influences fish physiology and can occasionally produce severe and lethal outcomes. Manganese exposure induces oxidative stress in Carassius auratus [103]. Manganese (Mn) contributes to numerous neurogenetic disorders by promoting the generation of free radicals and the inactivation of certain enzymes linked to antioxidant capacity [104]. Moreover, manganese adversely affects the liver and promotes cellular death in grouper [105].

Nichrome is widely utilized in several industrial processes and is regarded as a principal pollutant of aquatic ecosystems. In aquatic ecosystems, nickel (Ni) interacts with many chemical components to



E-ISSN: 2229-7677 • Website: www.ijsat.org • Email: editor@ijsat.org

create soluble salts that can adsorb onto other molecules, resulting in multiple synergistic and antagonistic effects [106]. The severity of nickel toxicity is contingent upon several elements, including nickel content, water quality, and the physiological condition of organisms [107]. Numerous investigations demonstrated that nickel accumulated in various organs of fish, particularly in the gills, leading to complications in respiratory processes [108, 109, 110, 111, 112]. Furthermore, Ni was observed to accumulate in the intestinal tract of fish, impairing its function [113, 114]. Nickel disrupts normal physiology and results in the mortality of certain freshwater fish species [115]. Nickel pollution causes various histological alterations in the gills of Orechromis niloticus, including hyperplasia, hypertrophy, and fusion of gill lamellae [116]. Moreover, Ni toxicity disrupts ion control [117, 118, 119] and provokes oxidative stress in fish [120, 121, 122, 123, 124]. Two investigations found no substantial effects on fish growth [125, 126]; however, they demonstrated significant affects on the growth of pulmonate snails [127] and zebrafish [128].

Lead is a very toxic metal that bioaccumulates in aquatic species via water and food sources [129]. Lead is bioaccumulated in various fish organs, including the liver, kidneys, gills, spleen, and digestive system [130, 131, 132, 133, 134, 135, 136]. Lead substantially alters the hematological parameters of fish [137, 138, 139, 140, 141]. Furthermore, Pb toxicity leads to a marked modification in enzyme activity within the blood plasma and liver of fish, resulting in various diseases in the cell membrane and damaging liver cells. Lead adversely impacts fish growth and feed efficiency by diminishing weight gain, specific growth rate, and feed consumption [142, 143, 144, 145]. Furthermore, lead (Pb) adversely affects reproductive performance, resulting in subpar sperm and ovum quality, diminished fertilization and hatching rates, and decreased survival of embryos and larvae.

Zn is a crucial micronutrient that significantly influences the growth and reproduction of fish; nevertheless, excessive Zn levels can have detrimental effects on fish. Zn pollution in aquatic habitats is well-documented [146, 147]. The liver and renal tissues are the primary locations for zinc bioaccumulation [148]. Zn poisoning adversely impacts the growth [149, 150, 151], homeostasis [152], feed consumption [153, 154, 155], and osseous development of fish [156]. Zinc poisoning prompts ammonia excretion, leading to deteriorated water quality and stressful environments for fish. Furthermore, zinc poisoning adversely affects the hepatic tissue of fish by elevating the activities of alanine aminotransferase (ALT) and aspartate aminotransferase (AST) [157, 158, 159]. Furthermore, elevated Zn concentrations markedly diminish the protein and fat content in fish, potentially leading to the oxidation of these macromolecules and inadequate protein consumption [160, 161].

The bioaccumulation of heavy metals in freshwater fish has significant ecological, environmental, and social ramifications. When metals are present at elevated concentrations in the environment, species acquire greater quantities. Inducing biomagnification of metals within the trophic web, adversely affecting aquatic ecology, which depends on these metals in multiple direct and indirect manners [162, 163].



E-ISSN: 2229-7677 • Website: www.ijsat.org • Email: editor@ijsat.org

5.0 Public Health Implications

Heavy metals are transferred through the food chain via many pathways, primarily originating from anthropogenic and natural sources [164]. Plants are crucial to this transfer process as they employ specific transport mechanisms to absorb heavy metals from the soil. Plants that accumulate heavy metals may undergo alterations in physiological and biochemical processes that impair their growth and development. Additionally, plants cultivated in soil contaminated with heavy metals may have nutrient deficiencies as a result of the adverse effects of heavy metals on plant nutrition. Due to the potential for heavy metals to bioaccumulate in food sources, this contamination presents significant health hazards [165].

Conversely, HM can be transported via air, water, or diverse species to numerous nearby sites, including rivers, ponds, lakes, and oceans. Aquatic creatures, especially fish, that bioaccumulate toxins subsequently expose other animals, including humans, through the food chain. A 2023 investigation in the Gulf of Guinea indicated the concentrations of heavy metals (HMs) in Penaeus notialis and D. angolensis, namely Cu: $12.08 \pm 1.46 \,\mu\text{g/g}$, Zn: $19.20 \pm 2.27 \,\mu\text{g/g}$, As: $8.46 \pm 2.42 \,\mu\text{g/g}$, Cd: $0.03 \pm 0.01 \,\mu\text{g/g}$, and Hg: $0.14 \pm 0.03 \,\mu\text{g/g}$ in D. angolensis [166]. Mercury was found in significant concentrations in D. angolensis [166]. The cancer risk associated with the ingestion of P. notialis surpassed the $10-6 \,\mu\text{m}$ threshold for all age demographics, according to publications [166].

While fish serves as a superior source of nutrients, environmental factors affect its nutritional value. Small quantities of metals pose a greater worry due to their potential for bioaccumulation and non-biodegradability [167]. Heavy metal toxicity, which can damage vital organs including the brain, central nervous system (CNS), and blood, may arise from the consumption of fish sourced from contaminated aquatic habitats [167-169]. Prolonged exposure can also induce neurological, muscular, and physical disorders such as Parkinson's disease, Alzheimer's disease, and multiple sclerosis in people [170-172]. A multitude of individuals experience allergies and extended exposure to certain metals that may lead to cancer [173, 174]. Toxicity from chemically significant heavy metals can result in disease, diminished quality of life, and ultimately death [175]. Due to their persistence, non-biodegradability, and capacity to accumulate in organs and tissues over time, heavy metals constitute a continual and non-biodegradable issue.

Heavy metals and metalloids are common environmental pollutants present in both aquatic and terrestrial environments [176, 177]. Their toxicity, bioaccumulation potential, and persistence all affect their level of hazard. The heavy metals and metalloids (Cr, Ni, Cu, Zn, Cd, Pb, Hg, and As) present the most significant hazards. The trophic transmission of these components in food chains significantly affects animal and population health. The concentrations of toxic heavy metals and metalloids in diverse environmental elements and local biota must be assessed and monitored. It is imperative to address hazardous heavy metals and metalloids that might cause significant harm to humans prior to their use, to alleviate their detrimental effects on human health and the environment.

6.0 Mitigation Strategies and Policy Recommendations

Bioremediation is an effective and environmentally benign method for restoring polluted ecosystems by eliminating harmful metals. Bioremediation of toxicants can be achieved through adsorption [178, 179, 180], physio-biochemical mechanisms [181, 182, 183, 184], and molecular mechanisms [185, 186, 187]. Various enzymes (superoxide dismutase, SOD; catalase, CAT; glutathione S transferase, GST) and



E-ISSN: 2229-7677 • Website: www.ijsat.org • Email: editor@ijsat.org

nonenzymatic substances (reduced glutathione, GSH) are pivotal in maintaining the balance of reactive oxygen species (ROS) through detoxification. SOD may convert superoxide radicals into hydrogen peroxide, which is then transformed into non-toxic oxygen and water by CAT enzymes [188]. Conversely, GST neutralizes toxicants by catalyzing electrophiles to glutathione (GSH). Furthermore, GSH undergoes nonenzymatic oxidation to form glutathione disulfide when interacting with electrophilic molecules, such as free radicals and reactive oxygen species (ROS).

Phytoremediation is a widely utilized bioremediation method that employs diverse plants and microorganisms to mitigate pollution in aquatic ecosystems. Microbial enzymes are crucial in transforming harmful pollutants into benign substances by modifying their chemical structure. Certain Lactobacillus species effectively remediate heavy metals by acidifying the environment and by biosorption or by establishing connections between heavy metals and their cellular components. Microorganisms exhibit resistance to heavy metals through several processes, including extracellular sequestration, internal sequestration, reduction of heavy metal ions by the microbial cell, and the formation of extracellular barriers.

A diverse array of microorganisms, including bacteria, fungus, and algae species, have been employed to detoxify heavy metals and maintain environmental cleanliness. Alongside natural microbes, many genetically modified microorganisms, particularly those with surface engineering, have been produced for the remediation of certain heavy metals [189]. Numerous studies indicate that genetically modified microorganisms possess superior capabilities compared to wild microbes in the removal of organic substances, including heavy metals, within natural environmental systems [190, 191].

Various engineering techniques, such as single-gene editing, metabolic pathway modification, and gene sequence alteration (both coding and regulatory), are effectively utilized to alter the genetic composition of microorganisms, thereby converting them into engineered microorganisms that more efficiently eliminate several heavy metals, including Ni, Hg, Cd, Fe, As, and Cu. The utilization of modern engineering methodologies (genomics, metagenomics, proteomics, metabolomics, and transcriptomics) has resulted in genetically engineered bacteria that are essential for the bioremediation of various heavy metals [192].

The utilization of genetically modified Pseudomonas putida and Escherichia coli strain M109 has effectively eliminated Hg from polluted locations [193]. The introduction of mer genes into Deinococcus geothemalis and Cupriavidus metallidurans strain MSR33 [194] has demonstrated effective mercury reduction. Furthermore, transporters in microbial membranes substantially enhance the bioremediation of heavy metals from the environment. Research has demonstrated that dietary Lactobacillus plantarum mitigates aluminum (Al) toxicity in tilapia [195]. Furthermore, dietary probiotics have been shown to mitigate the adverse effects of cadmium on the growth and hematological parameters of Oreochromis niloticus [196]. EDTA markedly diminished the body cadmium concentration, hence enhancing the blood profile of Clarias gariepinus [197].

Therefore, mitigating the environmental and public health impacts of heavy metal contamination in coastal and freshwater edible species requires a multifaceted and collaborative approach that integrates scientific monitoring, regulatory enforcement, and community participation. The first critical step involves strengthening environmental monitoring and surveillance systems. Regular assessment of water, sediment, and aquatic species using advanced analytical tools such as ICP-MS and AAS can help detect contamination trends and identify pollution hotspots early. Establishing a centralized database for heavy



E-ISSN: 2229-7677 • Website: www.ijsat.org • Email: editor@ijsat.org

metal monitoring would enable policymakers, researchers, and environmental agencies to track exposure levels and evaluate intervention outcomes more effectively.

Also, controlling pollution at the source is essential to prevent heavy metals from entering aquatic ecosystems. Governments should enforce strict regulations on industrial effluents, mining operations, and agricultural runoff, requiring industries to adopt cleaner production technologies and implement waste treatment before discharge. It is important to encourage the use of environmentally friendly alternatives to metal-based fertilizers and pesticides can also minimize diffuse pollution. In coastal regions, integrating pollution control into watershed management and urban planning will further reduce contamination from multiple sources.

From a policy standpoint, updating and harmonizing environmental and food safety standards with international guidelines such as those established by the WHO, FAO, and EPA—is necessary to ensure consistent protection levels. Policies should mandate routine inspection of seafood and freshwater fish sold in markets, with clear labeling of contamination risk levels to inform consumers. Public health authorities must develop and communicate consumption advisories, particularly for vulnerable populations such as children, pregnant women, and communities that rely heavily on fish as a dietary staple. Lastly, community education and stakeholder engagement play a vital role in sustaining long-term mitigation. Raising awareness about the sources and health effects of heavy metal contamination can empower local populations to advocate for cleaner environments and adopt safer fishing and consumption habits. Partnerships among governments, academia, NGOs, and the private sector should be fostered to support research, capacity building, and technology transfer. Through coordinated policy actions, scientific innovation, and public involvement, it is possible to significantly reduce heavy metal exposure and protect both aquatic ecosystems and human health.

Conclusion

Heavy metal contamination in coastal and freshwater ecosystems presents a serious threat to environmental sustainability, food security, and public health. The persistence and bioaccumulative nature of metals such as mercury, cadmium, lead, arsenic, and chromium enable them to infiltrate aquatic food webs, ultimately reaching humans through the consumption of contaminated fish and other edible species. The resulting health risks ranging from neurological and renal damage to developmental disorders and cancer explain the urgency of addressing this global issue through science-based interventions and robust policy actions. The findings of this study highlight that mitigating heavy metal pollution requires a holistic approach that connects environmental management with public health protection. Strengthening pollution control at industrial and agricultural sources, enforcing strict regulatory standards, and enhancing environmental monitoring are essential to reducing contamination levels. Equally important are ecosystem restoration initiatives and sustainable aquaculture practices that minimize bioaccumulation and ensure the long-term health of aquatic habitats. Ultimately, safeguarding human and ecological well-being demands collective responsibility among governments, researchers, industries, and communities to ensure cleaner water systems and safer food sources. By integrating scientific evidence into policy and promoting public awareness, societies can build resilient frameworks that not only reduce heavy metal exposure but also protect future generations from its enduring consequences.



E-ISSN: 2229-7677 • Website: www.ijsat.org • Email: editor@ijsat.org

References

- 1. Pal M, Ayele Y, Hadush M, Panigrahi S, Jadhav VJ. Public health hazards due to unsafe drinking water. Air Water Borne Dis. 2018 Jun;7(1000138):2.
- 2. Saxena V. Water quality, air pollution, and climate change: investigating the environmental impacts of industrialization and urbanization. Water, Air, & Soil Pollution. 2025 Feb;236(2):73.
- 3. Crain CM, Halpern BS, Beck MW, Kappel CV. Understanding and managing human threats to the coastal marine environment. Annals of the new York Academy of Sciences. 2009 Apr;1162(1):39-62.
- 4. Edo GI, Samuel PO, Oloni GO, Ezekiel GO, Ikpekoro VO, Obasohan P, Ongulu J, Otunuya CF, Opiti AR, Ajakaye RS, Essaghah AE. Environmental persistence, bioaccumulation, and ecotoxicology of heavy metals. Chemistry and Ecology. 2024 Mar 15;40(3):322-49.
- 5. Ali H, Khan E. Trophic transfer, bioaccumulation, and biomagnification of non-essential hazardous heavy metals and metalloids in food chains/webs—Concepts and implications for wildlife and human health. Human and Ecological Risk Assessment: An International Journal. 2019 Aug 18;25(6):1353-76.
- 6. Nkwunonwo UC, Odika PO, Onyia NI. A review of the health implications of heavy metals in food chain in Nigeria. The Scientific World Journal. 2020;2020(1):6594109.
- 7. Shah SB. Heavy metals in the marine environment—an overview. Heavy metals in Scleractinian corals. 2021 Apr 24:1-26.
- 8. Angon PB, Islam MS, Das A, Anjum N, Poudel A, Suchi SA. Sources, effects and present perspectives of heavy metals contamination: Soil, plants and human food chain. Heliyon. 2024 Apr 15;10(7).
- 9. Raj D, Maiti SK. Sources, bioaccumulation, health risks and remediation of potentially toxic metal (loid) s (As, Cd, Cr, Pb and Hg): an epitomised review. Environmental monitoring and assessment. 2020 Feb;192(2):108.
- 10. Jakimska-Nagórska A, Konieczka P, Skóra K, Namieśnik J. Bioaccumulation of metals in tissues of marine animals, part I: the role and impact of heavy metals on organisms. Polish Journal of Environmental Studies. 2011;20:1117-25.
- 11. Pandiyan J, Mahboob S, Govindarajan M, Al-Ghanim KA, Ahmed Z, Al-Mulhm N, Jagadheesan R, Krishnappa K. An assessment of level of heavy metals pollution in the water, sediment and aquatic organisms: A perspective of tackling environmental threats for food security. Saudi Journal of Biological Sciences. 2021 Feb 1;28(2):1218-25.
- 12. Tao Y, Yuan Z, Xiaona H, Wei M. Distribution and bioaccumulation of heavy metals in aquatic organisms of different trophic levels and potential health risk assessment from Taihu lake, China. Ecotoxicology and environmental safety. 2012 Jul 1;81:55-64.
- 13. Jiang X, Wang J, Pan B, Li D, Wang Y, Liu X. Assessment of heavy metal accumulation in freshwater fish of Dongting Lake, China: Effects of feeding habits, habitat preferences and body size. Journal of Environmental Sciences. 2022 Feb 1;112:355-65.
- 14. Rahman Z, Singh VP. The relative impact of toxic heavy metals (THMs)(arsenic (As), cadmium (Cd), chromium (Cr)(VI), mercury (Hg), and lead (Pb)) on the total environment: an overview. Environmental monitoring and assessment. 2019 Jul;191(7):419.
- 15. Sardar K, Ali S, Hameed S, Afzal S, Fatima S, Shakoor MB, Bharwana SA, Tauqeer HM. Heavy metals contamination and what are the impacts on living organisms. Greener Journal of Environmental management and public safety. 2013 Aug;2(4):172-9.



E-ISSN: 2229-7677 • Website: www.ijsat.org • Email: editor@ijsat.org

- 16. Singh A, Sharma A, Verma RK, Chopade RL, Pandit PP, Nagar V, Aseri V, Choudhary SK, Awasthi G, Awasthi KK, Sankhla MS. Heavy metal contamination of water and their toxic effect on living organisms. In The toxicity of environmental pollutants 2022 Jun 15. IntechOpen.
- 17. Aslam H, Umar A, Khan MU, Honey S, Ullah A, Ashraf MA, Ayesha G, Nusrat N, Jamil M, Khan S, Abid A. A Review on Heavy Metals in Ecosystems, their Sources, Roles, and Impact on Plant Life. Journal of Genetic Medicine. 2024;2994:256X.
- 18. Verma R, Dwivedi P. Heavy metal water pollution-A case study. Recent research in Science and Technology. 2013 Jul 27;5(5).
- 19. Teschke R. Aluminum, arsenic, beryllium, cadmium, chromium, cobalt, copper, iron, lead, mercury, molybdenum, nickel, platinum, thallium, titanium, vanadium, and zinc: molecular aspects in experimental liver injury. International journal of molecular sciences. 2022 Oct 13;23(20):12213.
- 20. Sharma DK, Nathan RJ, Kumar R, Jain AK. Heavy metal toxicity: impact on human health: a review. Indian J. Forensic Med. Pathol. 2021;14:270-8.
- 21. Shahjahan M., Taslima K., Rahman M.S., Al-Emran M., Alam S.I., Faggio C. Effects of Heavy Metals on Fish Physiology—A Review. Chemosphere. 2022;300:134519.
- 22. Taslima K., Al-Emran M., Rahman M.S., Hasan J., Ferdous Z., Rohani M.F., Shahjahan M. Impacts of Heavy Metals on Early Development, Growth and Reproduction of Fish—A Review. Toxicol. Rep. 2022;9:858–868.
- 23. Dixit R., Malaviya D., Pandiyan K., Singh U.B., Sahu A., Shukla R., Singh B.P., Rai J.P., Sharma P.K., Lade H., et al. Bioremediation of Heavy Metals from Soil and Aquatic Environment: An Overview of Principles and Criteria of Fundamental Processes. Sustainability. 2015;7:2189–2212.
- 24. Singare P.U., Dhabarde S.S. Toxic Metals Pollution Due to Industrial Effluents Released along Dombivali Industrial Belt of Mumbai, India. Eur. J. Environ. Saf. Sci. Eur. Sci. Res. Inst. 2014;2:5–11.
- 25. Sarkar T., Alam M.M., Parvin N., Fardous Z., Chowdhury A.Z., Hossain S., Haque M.E., Biswas N. Assessment of Heavy Metals Contamination and Human Health Risk in Shrimp Collected from Different Farms and Rivers at Khulna-Satkhira Region, Bangladesh. Toxicol. Rep. 2016;3:346–350.
- 26. Ezemonye L.I., Adebayo P.O., Enuneku A.A., Tongo I., Ogbomida E. Potential Health Risk Consequences of Heavy Metal Concentrations in Surface Water, Shrimp (Macrobrachium macrobrachion) and Fish (Brycinus longipinnis) from Benin River, Nigeria. Toxicol. Rep. 2019;6:1–9.
- 27. Authman M.M. Use of Fish as Bio-Indicator of the Effects of Heavy Metals Pollution. J. Aquac. Res. Dev. 2015;6:1–13.
- 28. Malik D.S., Maurya P.K. Heavy Metal Concentration in Water, Sediment, and Tissues of Fish Species (Heteropneustis fossilis and Puntius ticto) from Kali River, India. Toxicol. Environ. Chem. 2014;96:1195–1206.
- 29. Youssef D.H., Tayel F.T. Metal Accumulation by Three Tilapia Spp. From Some Egyptian Inland Waters. Chem. Ecol. 2004;20:61–71.
- 30. Farombi E.O., Adelowo O.A., Ajimoko Y.R. Biomarkers of Oxidative Stress and Heavy Metal Levels as Indicators of Environmental Pollution in African Cat Fish (Clarias gariepinus) from Nigeria Ogun River. Int. J. Environ. Res. Public Health. 2007;4:158–165.
- 31. Lu Y., Song S., Wang R., Liu Z., Meng J., Sweetman A.J., Jenkins A., Ferrier R.C., Li H., Luo W., et al. Impacts of Soil and Water Pollution on Food Safety and Health Risks in China. Environ. Int. 2015;77:5–15.



E-ISSN: 2229-7677 • Website: www.ijsat.org • Email: editor@ijsat.org

- 32. Gárriz Á., del Fresno P.S., Carriquiriborde P., Miranda L.A. Effects of Heavy Metals Identified in Chascomús Shallow Lake on the Endocrine-Reproductive Axis of Pejerrey Fish (Odontesthes bonariensis) Gen. Comp. Endocrinol. 2019;273:152–162.
- 33. Yan W., Hamid N., Deng S., Jia P.P., Pei D.S. Individual and Combined Toxicogenetic Effects of Microplastics and Heavy Metals (Cd, Pb, and Zn) Perturb Gut Microbiota Homeostasis and Gonadal Development in Marine Medaka (Oryzias melastigma) J. Hazard. Mater. 2020;397:122795.
- 34. Forouhar Vajargah M., Mohamadi Yalsuyi A., Sattari M., Prokić M.D., Faggio C. Effects of Copper Oxide Nanoparticles (CuO-NPs) on Parturition Time, Survival Rate and Reproductive Success of Guppy Fish. Poecilia reticulata. J. Clust. Sci. 2020;31:499–506.
- 35. Cao L., Huang W., Shan X., Xiao Z., Wang Q., Dou S. Cadmium Toxicity to Embryonic-Larval Development and Survival in Red Sea Bream Pagrus major. Ecotoxicol. Environ. Saf. 2009;72:1966–1974.
- 36. Siddiquee S., Rovina K., Azad S. Al Heavy Metal Contaminants Removal from Wastewater Using the Potential Filamentous Fungi Biomass: A Review. J. Microb. Biochem. Technol. 2015;7:384–395.
- 37. Maurya P.K., Malik D.S., Yadav K.K., Kumar A., Kumar S., Kamyab H. Bioaccumulation and Potential Sources of Heavy Metal Contamination in Fish Species in River Ganga Basin: Possible Human Health Risks Evaluation. Toxicol. Rep. 2019;6:472–481.
- 38. Ahluwalia S.S., Goyal D. Microbial and Plant Derived Biomass for Removal of Heavy Metals from Wastewater. Bioresour. Technol. 2007;98:2243–2257.
- 39. Igiri B.E., Okoduwa S.I.R., Idoko G.O., Akabuogu E.P., Adeyi A.O., Ejiogu I.K. Toxicity and Bioremediation of Heavy Metals Contaminated Ecosystem from Tannery Wastewater: A Review. J. Toxicol. 2018;2018:1–16.
- 40. Akcil A., Erust C., Ozdemiroglu S., Fonti V., Beolchini F. A Review of Approaches and Techniques Used in Aquatic Contaminated Sediments: Metal Removal and Stabilization by Chemical and Biotechnological Processes. J. Clean. Prod. 2015;86:24–36.
- 41. Velma V., Vutukuru S.S., Tchounwou P.B. Ecotoxicology of Hexavalent Chromium in Freshwater Fish: A Critical Review. Rev. Environ. Health. 2009;24:129–145.
- 42. Fernandes C., Fontaínhas-Fernandes A., Cabral D., Salgado M.A. Heavy Metals in Water, Sediment and Tissues of Liza saliens from Esmoriz-Paramos Lagoon, Portugal. Environ. Monit. Assess. 2008;136:267–275.
- 43. Al-Busaidi M., Yesudhason P., Al-Mughairi S., Al-Rahbi W.A.K., Al-Harthy K.S., Al-Mazrooei N.A., Al-Habsi S.H. Toxic Metals in Commercial Marine Fish in Oman with Reference to National and International Standards. Chemosphere. 2011;85:67–73.
- 44. El-Moselhy K.M., Othman A.I., Abd El-Azem H., El-Metwally M.E.A. Bioaccumulation of Heavy Metals in Some Tissues of Fish in the Red Sea, Egypt. Egypt. J. Basic Appl. Sci. 2014;1:97–105.
- 45. Mokarram M., Saber A., Sheykhi V. Effects of Heavy Metal Contamination on River Water Quality Due to Release of Industrial Effluents. J. Clean. Prod. 2020;277:123380.
- 46. Yu B., Wang X., Dong K.F., Xiao G., Ma D. Heavy Metal Concentrations in Aquatic Organisms (Fishes, Shrimp and Crabs) and Health Risk Assessment in China. Mar. Pollut. Bull. 2020;159:111505.
- 47. Garelic H., Jones H., Dybowska A., Valsami-Jones E. Arsenic Pollution Sources. Rev. Environ. Contam. 2009;197:17–60.



- 48. Suhendrayatna, Ohki A., Nakajima T., Maeda S. Studies on the Accumulation and Transformation of Arsenic in Freshwater Organisms II. Accumulation and Transformation of Arsenic Compounds by Tilapia mossambica. Chemosphere. 2002;46:325–331.
- 49. Hayati A., Giarti K., Winarsih Y., Amin M.H.F. The Effect of Cadmium on Sperm Quality and Fertilization Of Cyprinus carpio L. J. Trop. Biodivers. Biotechnol. 2017;2:45.
- 50. Zhang W., Chen L., Zhou Y., Wu Y., Zhang L. Biotransformation of Inorganic Arsenic in a Marine Herbivorous Fish Siganus Fuscescens after Dietborne Exposure. Chemosphere. 2016;147:297–304.
- 51. Pei J., Zuo J., Wang X., Yin J., Liu L., Fan W. The Bioaccumulation and Tissue Distribution of Arsenic Species in Tilapia. Int. J. Environ. Res. Public Health. 2019;16:757.
- 52. Wiszniewska A., Hanus-Fajerska E., MuszyŃska E., Ciarkowska K. Natural Organic Amendments for Improved Phytoremediation of Polluted Soils: A Review of Recent Progress. Pedosphere. 2016;26:1–12.
- 53. Baker D.E., Chesnin L. Chemical Monitoring of Soils for Environmental Quality and Animal and Human Health. Adv. Agron. 2002;27:305–374.
- 54. Kapaj S., Peterson H., Liber K., Bhattacharya P. Human Health Effects from Chronic Arsenic Poisoning—A Review. J. Environ. Sci. Health-Part A Toxic/Hazard. Subst. Environ. Eng. 2006;41:2399–2428.
- 55. Erickson R.J., Mount D.R., Highland T.L., Russell Hockett J., Jenson C.T. The Relative Importance of Waterborne and Dietborne Arsenic Exposure on Survival and Growth of Juvenile Rainbow Trout. Aquat. Toxicol. 2011;104:108–115.
- 56. Sayed A.E.D.H., Elbaghdady H.A.M., Zahran E. Arsenic-Induced Genotoxicity in Nile Tilapia (Orechromis niloticus); The Role of Spirulina Platensis Extract. Environ. Monit. Assess. 2015;187:1–10.
- 57. Kumari B., Kumar V., Sinha A.K., Ahsan J., Ghosh A.K., Wang H., DeBoeck G. Toxicology of Arsenic in Fish and Aquatic Systems. Environ. Chem. Lett. 2017;15:43–64.
- 58. Ghosh D., Bhattacharya S., Mazumder S. Perturbations in the Catfish Immune Responses by Arsenic: Organ and Cell Specific Effects. Comp. Biochem. Physiol.-C Toxicol. Pharmacol. 2006;143:455–463.
- 59. Nayak A.S., Lage C.R., Kim C.H. Effects of Low Concentrations of Arsenic on the Innate Immune System of the Zebrafish (Danio rerio) Toxicol. Sci. 2007;98:118–124.
- 60. Datta S., Ghosh D., Saha D.R., Bhattacharaya S., Mazumder S. Chronic Exposure to Low Concentration of Arsenic Is Immunotoxic to Fish: Role of Head Kidney Macrophages as Biomarkers of Arsenic Toxicity to Clarias batrachus. Aquat. Toxicol. 2009;92:86–94.
- 61. Fazio F., Saoca C., Ferrantelli V., Cammilleri G., Capillo G., Piccione G. Relationship between Arsenic Accumulation in Tissues and Hematological Parameters in Mullet Caught in Faro Lake: A Preliminary Study. Environ. Sci. Pollut. Res. 2019;26:8821–8827.
- 62. Akter M.S. Acute Toxicity of Arsenic and Mercury to Fresh Water Climbing Perch, Anabas testudineus (Bloch) World J. Zool. 2008;3:13–18.
- 63. Baldissarelli L.A., Capiotti K.M., Bogo M.R., Ghisleni G., Bonan C.D. Arsenic Alters Behavioral Parameters and Brain Ectonucleotidases Activities in Zebrafish (Danio rerio) Comp. Biochem. Physiol.-C Toxicol. Pharmacol. 2012;155:566–572.
- 64. Sayed A.H., Kitamura D., Oda S., Kashiwada S., Mitani H. Cytotoxic and Genotoxic Effects of Arsenic on Erythrocytes of Oryzias latipes: Bioremediation Using Spirulina Platensis. J. Trace Elem. Med. Biol. 2019;55:82–88.



- 65. Yamaguchi S., Miura C., Ito A., Agusa T., Iwata H., Tanabe S., Tuyen B.C., Miura T. Effects of Lead, Molybdenum, Rubidium, Arsenic and Organochlorines on Spermatogenesis in Fish: Monitoring at Mekong Delta Area and in Vitro Experiment. Aquat. Toxicol. 2007;83:43–51.
- 66. Boyle D., Brix K.V., Amlund H., Lundebye A.K., Hogstrand C., Bury N.R. Natural Arsenic Contaminated Diets Perturb Reproduction in Fish. Environ. Sci. Technol. 2008;42:5354–5360.
- 67. Celino F.T., Yamaguchi S., Miura C., Miura T. Arsenic Inhibits In Vitro Spermatogenesis and Induces Germ Cell Apoptosis in Japanese Eel (Anguilla japonica) Reproduction. 2009;138:279–287.
- 68. Zhou Q., Zhang J., Fu J., Shi J., Jiang G. Biomonitoring: An Appealing Tool for Assessment of Metal Pollution in the Aquatic Ecosystem. Anal. Chim. Acta. 2008;606:135–150.
- 69. Taweel A., Shuhaimi-Othman M., Ahmad A.K. Assessment of Heavy Metals in Tilapia Fish (Oreochromis niloticus) from the Langat River and Engineering Lake in Bangi, Malaysia, and Evaluation of the Health Risk from Tilapia Consumption. Ecotoxicol. Environ. Saf. 2013;93:45–51.
- 70. Chahid A., Hilali M., Benlhachimi A., Bouzid T. Contents of Cadmium, Mercury and Lead in Fish from the Atlantic Sea (Morocco) Determined by Atomic Absorption Spectrometry. Food Chem. 2014;147:357–360.
- 71. Reynders H., Van Campenhout K., Bervoets L., De Coen W.M., Blust R. Dynamics of Cadmium Accumulation and Effects in Common Carp (Cyprinus carpio) during Simultaneous Exposure to Water and Food (Tubifex Tubifex) Environ. Toxicol. Chem. 2006;25:1558–1567.
- 72. ratap H.B. Effects of Ambient and Dietary Cadmium on Haematological Parametres in Oreochromis mossambicus Acclimatised to Low- and High-Calcium Water. Comp. Clin. Path. 2008;17:133–136.
- 73. Almeida J.A., Diniz Y.S., Marques S.F.G., Faine L.A., Ribas B.O., Burneiko R.C., Novelli E.L.B. The Use of the Oxidative Stress Responses as Biomarkers in Nile Tilapia (Oreochromis niloticus) Exposed to in Vivo Cadmium Contamination. Environ. Int. 2002;27:673–679.
- 74. Valavanidis A., Vlahogianni T., Dassenakis M., Scoullos M. Molecular Biomarkers of Oxidative Stress in Aquatic Organisms in Relation to Toxic Environmental Pollutants. Ecotoxicol. Environ. Saf. 2006;64:178–189.
- 75. Li Z.H., Li P., Dzyuba B., Randak T. Influence of Environmental Related Concentrations of Heavy Metals on Motility Parameters and Antioxidant Responses in Sturgeon Sperm. Chem. Biol. Interact. 2010;188:473–477.
- 76. Dietrich G.J., Dietrich M., Kowalski R.K., Dobosz S., Karol H., Demianowicz W., Glogowski J. Exposure of Rainbow Trout Milt to Mercury and Cadmium Alters Sperm Motility Parameters and Reproductive Success. Aquat. Toxicol. 2010;97:277–284.
- 77. Dietrich G.J., Ciereszko A., Kowalski R.K., Rzemieniecki A., Bogdan E., Demianowicz W., Dietrich M., Kujawa R., Glogowski J. Motility and Fertilizing Capacity of Frozen/Thawed Sperm of Siberian Sturgeon after a Short-Time Exposure of Fresh Semen to Mercury and Cadmium. J. Appl. Ichthyol. 2012;28:973–977.
- 78. Monteiro M.I.C., Fraga I.C.S., Yallouz A.V., De Oliveira N.M.M., Ribeiro S.H. Determination of Total Chromium Traces in Tannery Effluents by Electrothermal Atomic Absorption Spectrometry, Flame Atomic Absorption Spectrometry and UV-Visible Spectrophotometric Methods. Talanta. 2002;58:629–633.
- 79. Palaniappan P.R., Karthikeyan S. Bioaccumulation and Depuration of Chromium in the Selected Organs and Whole Body Tissues of Freshwater Fish Cirrhinus mrigala Individually and in Binary Solutions with Nickel. J. Environ. Sci. 2009;21:229–236.



- 80. Vinodhini R., Narayanan M. Bioaccumulation of Heavy Metals in Organs of Fresh Water Fish Cyprinus carpio (Common Carp) Int. J. Environ. Sci. Technol. 2008;5:179–182.
- 81. Ali Z., Yousafzai A.M., Sher N., Muhammad I., Nayab G.E., Aqeel S.A.M., Shah S.T., Aschner M., Khan I., Khan H. Toxicity and Bioaccumulation of Manganese and Chromium in Different Organs of Common Carp (Cyprinus carpio) Fish. Toxicol. Rep. 2021;8:343–348.
- 82. Fawad M., Yousafzai A.M., Haseeb A., Rehman H.U., Akhtar P.N., Saeed K., Khan University Mardan Buner Campus. Khalid Usman W., Muhammad Fawad C., jan Afridi A., et al. Acute Toxicity and Bioaccumulation of Chromium in Gills, Skin and Intestine of Goldfish (Carassius auratus) J. Entomol. Zool. Stud. 2017;5:568–571.
- 83. El Shafei H.M. Bioaccumulation of Hexavalent Chromium in Tissues of a Freshwater Fish. Biochem. Anal. Biochem. 2016;5:2–5. doi: 10.4172/2161-1009.1000272.
- 84. Shaheen T., Jabeen F. Effect of Various Doses of Cr (VI) on Survival and Growth of Cyprinus carpio. Pak. J. Zool. 2015;47:913–919.
- 85. Madhavan P., Elumalai K. Effects of Chromium (VI) on the Lipid Peroxidation and Antioxidant Parameters in the Gill and Kidney Tissues of Catfish, Clarias batrachus (Linnaeus, 1758) (Actinopterygii: Siluriformes) Int. J. Adv. Res. Biol. Sci. 2016;3:249–255.
- 86. Bakshi A., Panigrahi A.K. A Comprehensive Review on Chromium Induced Alterations in Fresh Water Fishes. Toxicol. Rep. 2018;5:440–447.
- 87. Islam S.M.M., Rohani M.F., Zabed S.A., Islam M.T., Jannat R., Akter Y., Shahjahan M. Acute Effects of Chromium on Hemato-Biochemical Parameters and Morphology of Erythrocytes in Striped Catfish Pangasianodon hypophthalmus. Toxicol. Rep. 2020;7:664–670.
- 88. Selcuk Z., Tiril S.U., Alagil F., Belen V., Salman M., Cenesiz S., Muglali O.H., Yagci F.B. Effects of Dietary L-Carnitine and Chromium Picolinate Supplementations on Performance and Some Serum Parameters in Rainbow Trout (Oncorhynchus mykiss) Aquac. Int. 2010;18:213–221.
- 89. Chen H., Cao J., Li L., Wu X., Bi R., Klerks P.L., Xie L. Maternal Transfer and Reproductive Effects of Cr(VI) in Japanese Medaka (Oryzias latipes) under Acute and Chronic Exposures. Aquat. Toxicol. 2016;171:59–68.
- 90. Ni X., Shen Y. Transgenerational Effects of Hexavalent Chromium on Marine Medaka (Oryzias melastigma) Reveal Complex Transgenerational Adaptation in Offspring. Biomolecules. 2021;11:138.
- 91. Mishra A.K., Mohanty B. Histopathological Effects of Hexavalent Chromium in the Ovary of a Fresh Water Fish, Channa punctatus (Bloch) Bull. Environ. Contam. Toxicol. 2008;80:507–511.
- 92. Singh V.K., Singh K.P., Mohan D. Status of Heavy Metals in Water and Bed Sediments of River Gomti-A Tributary of the Ganga River, India. Environ. Monit. Assess. 2005;105:43–67.
- 93. Paul D. Research on Heavy Metal Pollution of River Ganga: A Review. Ann. Agrar. Sci. 2017;15:278–286.
- 94. Mitra S., Sarkar S.K., Raja P., Biswas J.K., Murugan K. Dissolved Trace Elements in Hooghly (Ganges) River Estuary, India: Risk Assessment and Implications for Management. Mar. Pollut. Bull. 2018;133:402–414.
- 95. Çoğun H.Y., Yüzereroğlu T.A., Kargin F. Accumulation of Copper and Cadmium in Small and Large Nile Tilapia Oreochromis niloticus. Bull. Environ. Contam. Toxicol. 2003;71:1265–1271.
- 96. Subathra S., Karuppasamy R. Bioaccumulation and Depuration Pattern of Copper in Different Tissues of Mystus vittatus, Related to Various Size Groups. Arch. Environ. Contam. Toxicol. 2008;54:236–244.



- 97. Aldoghachi M.A.J., Rahman M.M., Yusoff I., Sofian-Azirun M. Acute Toxicity and Bioaccumulation of Heavy Metals in Red Tilapia Fish. J. Anim. Plant Sci. 2016;26:507–513.
- 98. Shukla V., Dhankhar M., Prakash J., Sastry K.V. Bioaccumulation of Zn, Cu and Cd in Channa punctatus. J. Environ. Biol. 2007;28:395–397.
- 99. Alsop D., Brown S., Van Der Kraak G. The Effects of Copper and Benzo[a]Pyrene on Retinoids and Reproduction in Zebrafish. Aquat. Toxicol. 2007;82:281–295.
- 100. Howe P.D., Malcolm M.H.M., Dobson D.S. Manganese and its Compounds: Environmental Aspects. World Health Organization; Geneva, Switzerland: 2005
- 101. Morillo J., Usero J. Trace Metal Bioavailability in the Waters of Two Different Habitats in Spain: Huelva Estuary and Algeciras Bay. Ecotoxicol. Environ. Saf. 2008;71:851–859.
- 102. Partridge G.J., Lymbery A.J. Effects of Manganese on Juvenile Mulloway (Argyrosomus Japonicus) Cultured in Water with Varying Salinity-Implications for Inland Mariculture. Aquaculture. 2009;290:311–316.
- 103. Vieira M.C., Torronteras R., Córdoba F., Canalejo A. Acute Toxicity of Manganese in Goldfish Carassius Auratus Is Associated with Oxidative Stress and Organ Specific Antioxidant Responses. Ecotoxicol. Environ. Saf. 2012;78:212–217.
- 104. Chen M.T., Cheng G.W., Lin C.C., Chen B.H., Huang Y.L. Effects of Acute Manganese Chloride Exposure on Lipid Peroxidation and Alteration of Trace Metals in Rat Brain. Biol. Trace Elem. Res. 2006;110:163–177.
- 105. Wang X., Liu B.L., Gao X.Q., Fang Y.Y., Zhang X.H., Cao S.Q., Zhao K.F., Wang F. Effect of Long-Term Manganese Exposure on Oxidative Stress, Liver Damage and Apoptosis in Grouper Epinephelus moara × Epinephelus lanceolatus. Front. Mar. Sci. 2022;9:1–11.
- 106. Eisler R. Nickel Hazards to Fish, Wildlife, and Invertebrates: A Synoptic Review, U.S. Geological Survey. Biol. Sci. Rep. 1998;34:95.
- 107. Pyle G., Couture P. Homeostasis and Toxicology of Essential Metals. In: Wood C.M., Farrell A.P., Brauner C.J., editors. Fish Physiology. Volume 31. Academic Press; Cambridge, MA, USA: 2012. pp. 253–289.
- 108. Pane E.F., Richards J.G., Wood C.M. Acute Waterborne Nickel Toxicity in the Rainbow Trout (Oncorhynchus mykiss) Occurs by a Respiratory Rather than Ionoregulatory Mechanism. Aquat. Toxicol. 2003;63:65–82.
- 109. Pane E.F., Haque A., Goss G.G., Wood C.M. The Physiological Consequences of Exposure to Chronic, Sufblethal Waterborne Nickel in Rainbow Trout (Oncorhynchus mykiss): Exercise vs Resting Physiology. J. Exp. Biol. 2004;207:1249–1261.
- 110. ane E.F., Haque A., Wood C.M. Mechanistic Analysis of Acute, Ni-Induced Respiratory Toxicity in the Rainbow Trout (Oncorhynchus mykiss): An Exclusively Branchial Phenomenon. Aquat. Toxicol. 2004;69:11–24.
- 111. Brix K.V., Keithly J., DeForest D.K., Laughlin J. Acute and Chronic Toxicity of Nickel to Rainbow Trout (Oncorhynchus mykiss) Environ. Toxicol. Chem. 2004;23:2221–2228.
- 112. Blewett T.A., Wood C.M., Glover C.N. Salinity-Dependent Nickel Accumulation and Effects on Respiration, Ion Regulation and Oxidative Stress in the Galaxiid Fish, Galaxias maculatus. Environ. Pollut. 2016;214:132–141.
- 113. Blewett T.A., Wood C.M. Salinity-Dependent Nickel Accumulation and Oxidative Stress Responses in the Euryhaline Killifish (Fundulus heteroclitus) Arch. Environ. Contam. Toxicol. 2015;68:382–394.



- 114. Blewett T.A., Ransberry V.E., McClelland G.B., Wood C.M. Investigating the Mechanisms of Ni Uptake and Sub-Lethal Toxicity in the Atlantic Killifish Fundulus Heteroclitus in Relation to Salinity. Environ. Pollut. 2016;211:370–381.
- 115. Svecevičius G. Acute Toxicity of Nickel to Five Species of Freshwater Fish. Polish J. Environ. Stud. 2010;19:453–456.
- 116. Al-Attar A.M. The Influences of Nickel Exposure on Selected Physiological Parameters and Gill Structure in the Teleost Fish, Oreochromis niloticus. J. Biol. Sci. 2007;7:77–85.
- 117. Leonard E.M., Barcarolli I., Silva K.R., Wasielesky W., Wood C.M., Bianchini A. The Effects of Salinity on Acute and Chronic Nickel Toxicity and Bioaccumulation in Two Euryhaline Crustaceans: Litopenaeus vannamei and Excirolana armata. Comp. Biochem. Physiol.-C Toxicol. Pharmacol. 2011;154:409–419.
- 118. Leonard E.M., Marentette J.R., Balshine S., Wood C.M. Critical Body Residues, Michaelis-Menten Analysis of Bioaccumulation, Lethality and Behaviour as Endpoints of Waterborne Ni Toxicity in Two Teleosts. Ecotoxicology. 2014;23:147–162.
- 119. Leonard E.M., Wood C.M. Acute Toxicity, Critical Body Residues, Michaelis-Menten Analysis of Bioaccumulation, and Ionoregulatory Disturbance in Response to Waterborne Nickel in Four Invertebrates: Chironomus riparius, Lymnaea stagnalis, Lumbriculus variegatus and Daphnia pulex. Comp. Biochem. Physiol.-C Toxicol. Pharmacol. 2013;158:10–21.
- 120. James R., Sampath K. Effect of Zeolite on the Reduction of Cadmium Toxicity in Water and a Freshwater Fish, Oreochromis mossambicus. Bull. Environ. Contam. Toxicol. 1999;62:222–229.
- 121. ubrak O.I., Rovenko B.M., Husak V.V., Storey J.M., Storey K.B., Lushchak V.I. Nickel Induces Hyperglycemia and Glycogenolysis and Affects the Antioxidant System in Liver and White Muscle of Goldfish Carassius auratus L. Ecotoxicol. Environ. Saf. 2012;80:231–237.
- 122. Kubrak O.I., Husak V.V., Rovenko B.M., Poigner H., Kriews M., Abele D., Lushchak V.I. Antioxidant System Efficiently Protects Goldfish Gills from Ni2+-Induced Oxidative Stress. Chemosphere. 2013;90:971–976.
- 123. Kubrak O.I., Poigner H., Husak V.V., Rovenko B.M., Meyer S., Abele D., Lushchak V.I. Goldfish Brain and Heart Are Well Protected from Ni2+-Induced Oxidative Stress. Comp. Biochem. Physiol. Part-C Toxicol. Pharmacol. 2014;162:43–50.
- 124. Blewett T.A., Leonard E.M. Mechanisms of Nickel Toxicity to Fish and Invertebrates in Marine and Estuarine Waters. Environ. Pollut. 2017;223:311–322.
- 125. Ptashynski M.D., Pedlar R.M., Evans R.E., Baron C.L., Klaverkamp J.F. Toxicology of Dietary Nickel in Lake Whitefish (Coregonus clupeaformis) Aquat. Toxicol. 2002;58:229–247.
- 126. Bielmyer G.K., Jarvis T.A., Harper B.T., Butler B., Rice L., Ryan S., McLoughlin P. Metal Accumulation from Dietary Exposure in the Sea Urchin, Strongylocentrotus droebachiensis. Arch. Environ. Contam. Toxicol. 2012;63:86–94.
- 127. Niyogi S., Brix K.V., Grosell M. Effects of Chronic Waterborne Nickel Exposure on Growth, Ion Homeostasis, Acid-Base Balance, and Nickel Uptake in the Freshwater Pulmonate Snail, Lymnaea stagnalis. Aquat. Toxicol. 2014;150:36–44.
- 128. Alsop D., Lall S.P., Wood C.M. Reproductive Impacts and Physiological Adaptations of Zebrafish to Elevated Dietary Nickel. Comp. Biochem. Physiol. Part-C Toxicol. Pharmacol. 2014;165:67–75.
- 129. Sfakianakis D.G., Renieri E., Kentouri M., Tsatsakis A.M. Effect of Heavy Metals on Fish Larvae Deformities: A Review. Environ. Res. 2015;137:246–255.



- 130. Jezierska B., Witeska M. The Metal Uptake and Accumulation in Fish Living in Polluted Waters Bt-Soil and Water Pollution Monitoring, Protection and Remediation. Springer; Berlin/Heidelberg, Germany: 2006. pp. 107–114.
- 131. Adeyemo O.K. Haematological Profile of Clarias gariepinus (Burchell, 1822) Exposed to Lead. Turkish J. Fish. Aquat. Sci. 2007;7:163–169.
- 132. Dai W., Du H., Fu L., Jin C., Xu Z., Liu H. Effects of Dietary Pb on Accumulation, Histopathology, and Digestive Enzyme Activities in the Digestive System of Tilapia (Oreochromis niloticus) Biol. Trace Elem. Res. 2009;127:124–131.
- 133. Victor K., Patience A. Accumulation of Lead in the Tissues of Freshwater Catfish Clarias Gariepinus Exposed to Static Nominal Concentrations of Lead Nitrate. Agric. Biol. J. N. Am. 2012;3:510–515.
- 134. Banaee M., Haghi B.N., Zoheiri F. Original Article LC 50 and Bioaccumulation of Lead Nitrate (Pb(NO3)2) in Goldfish (Carassius auratus) Int. J. Aquat. Biol. 2013;1:233–239.
- 135. Khalesi M.K., Abedi Z., Behrouzi S., Kohestan Eskandari S. Haematological, Blood Biochemical and Histopathological Effects of Sublethal Cadmium and Lead Concentrations in Common Carp. Bulg. J. Vet. Med. 2017;20:141–150.
- 136. bdel-Warith A.W.A., Younis E.S.M.I., Al-Asgah N.A., Rady A.M., Allam H.Y. Bioaccumulation of Lead Nitrate in Tissues and Its Effects on Hematological and Biochemical Parameters of Clarias gariepinus. Saudi J. Biol. Sci. 2020;27:840–845.
- 137. Shah S.L. Hematological Parameters in Tench Tinca Tinca after Short Term Exposure to Lead. J. Appl. Toxicol. 2006;26:223–228.
- 138. Tawari-Fufeyin P., Igetei J., Okoidigun M.E. Changes in the Catfish (Clarias gariepinus) Exposed to Acute Cadmium and Lead Poisoning. Biosci. Res. Commun. 2008;20:271–276.
- 139. Stanley O.N., Omerebele U.A.M. Changes in the Haematological Parameters of Clarias gariepinus Exposed to Lead Poisoning. J. Fish. Int. 2010;5:72–76.
- 140. Mahmoud U., Ebied A., Mohamed S. Effect of Lead on Some Haematological and Biochemical Characteristics of Clarias gariepinus Dietary Supplemented with Lycopene and Vitamin E. Egypt. Acad. J. Biol. Sci. C Physiol. Mol. Biol. 2013;5:67–89.
- 141. Ikeogu C.F., Nsofor C.I., Igwilo I.O., Ngene A.A. Haematological and Serological Responses of Clarias gariepinus to Sublethal Concentrations of Lead Nitrate. J. Pharm. Sci. Biosci. Res. 2016;6:442–446.
- 142. Yang J.L., Chen H.C. Serum Metabolic Enzyme Activities and Hepatocyte Ultrastructure of Common Carp after Gallium Exposure. Zool. Stud. 2003;42:455–461
- 143. Pérez-Rostro C.I., Racotta I.S., Ibarra A.M. Decreased Genetic Variation in Metabolic Variables of Litopenaeus vannamei Shrimp after Exposure to Acute Hypoxia. J. Exp. Mar. Bio. Ecol. 2004;302:189–200.
- 144. Olojo E.A.A., Abass A.A., Olurin K.B., Mbaka G. The Potential Use of Certain Protein Metabolism Parameters as Biomarkers of Heavy Metal (Lead) Stress in the African Catfish, Clarias gariepinus. Agric. J. 2012;7:316–322.
- 145. Javed M. Effects of Zinc and Lead Toxicity on the Growth and Their Bioaccumulation in Fish. Pak. Vet. J. 2012;32:357–362.
- 146. Senthil Murugan S., Karuppasamy R., Poongodi K., Puvaneswari S. Bioaccumulation Pattern of Zinc in Freshwater Fish Channa punctatus (Bloch.) after Chronic Exposure. Turkish J. Fish. Aquat. Sci. 2008;59:55–59.



- 147. Bowen L., Werner I., Johnson M.L. Physiological and Behavioral Effects of Zinc and Temperature on Coho Salmon (Oncorhynchus kisutch) Hydrobiologia. 2006;559:161–168. doi: 10.1007/s10750-005-1095-3.
- 148. Abdel-Tawwab M., El-Sayed G.O., Shady S.H. Growth, Biochemical Variables, and Zinc Bioaccumulation in Nile Tilapia, Oreochromis niloticus (L.) as Affected by Water-Born Zinc Toxicity and Exposure Period. Int. Aquat. Res. 2016;8:197–206.
- 149. hiau S.Y., Jiang L.C. Dietary Zinc Requirements of Grass Shrimp, Penaeus monodon, and Effects on Immune Responses. Aquaculture. 2006;254:476–482
- 150. Abdel-Tawwab M., El-Sayed G.O., Shady S.H. Effects of Dietary Protein Levels and Environmental Zinc Exposure on the Growth, Feed Utilization, and Biochemical Variables of Nile Tilapia, Oreochromis niloticus (L.) Toxicol. Environ. Chem. 2012;94:1368–1382.
- 151. Abdel-Tawwab M., Mousaad M.N., Sharafeldin K.M., Ismaiel N.E. Changes in Growth and Biochemical Status of Common Carp, Cyprinus carpio L. Exposed to Water-Born Zinc Toxicity for Different Periods. Int. Aquat. Res. 2013;5:11.
- 152. Xiong D., Fang T., Yu L., Sima X., Zhu W. Effects of Nano-Scale TiO2, ZnO and Their Bulk Counterparts on Zebrafish: Acute Toxicity, Oxidative Stress and Oxidative Damage. Sci. Total Environ. 2011;409:1444–1452.
- 153. armo e Sa M.V.d., Pezzato L.E., Ferreira Lima M.M.B., De Magalhães Padilha P. Optimum Zinc Supplementation Level in Nile Tilapia Oreochromis niloticus Juveniles Diets. Aquaculture. 2004;238:385–401.
- 154. Tan L.N., Feng L., Liu Y., Jiang J., Jiang W.D., Hu K., Li S.H., Zhou X.Q. Growth, Body Composition and Intestinal Enzyme Activities of Juvenile Jian Carp (Cyprinus carpio Var. Jian) Fed Graded Levels of Dietary Zinc. Aquac. Nutr. 2011;17:338–345.
- 155. Luo Z., Tan X.Y., Zheng J.L., Chen Q.L., Liu C.X. Quantitative Dietary Zinc Requirement of Juvenile Yellow Catfish Pelteobagrus fulvidraco, and Effects on Hepatic Intermediary Metabolism and Antioxidant Responses. Aquaculture. 2011;319:150–155.
- 156. Salvaggio A., Marino F., Albano M., Pecoraro R., Camiolo G., Tibullo D., Bramanti V., Lombardo B.M., Saccone S., Mazzei V., et al. Toxic Effects of Zinc Chloride on the Bone Development in Danio rerio (Hamilton, 1822) Front. Physiol. 2016;7:1–6.
- 157. Coppo J.A., Mussart N.B., Fioranelli S.A. Physiological Variation of Enzymatic Activities in Blood of Bullfrog, Rana Catesbeiana (Shaw, 1802) Rev. Vet. 2002;216:2001–2002.
- 158. Chen C.Y., Wooster G.A., Bowser P.R. Comparative Blood Chemistry and Histopathology of Tilapia Infected with Vibrio vulnificus or Streptococcus iniae or Exposed to Carbon Tetrachloride, Gentamicin, or Copper Sulfate. Aquaculture. 2004;239:421–443.
- 159. Firat Ö., Kargin F. Individual and Combined Effects of Heavy Metals on Serum Biochemistry of Nile Tilapia Oreochromis niloticus. Arch. Environ. Contam. Toxicol. 2010;58:151–157.
- 160. Cakmak G., Togan I., Severcan F. 17β-Estradiol Induced Compositional, Structural and Functional Changes in Rainbow Trout Liver, Revealed by FT-IR Spectroscopy: A Comparative Study with Nonylphenol. Aquat. Toxicol. 2006;77:53–63.
- 161. Abdel-Tawwab M., Khattab Y.A.E., Ahmad M.H., Shalaby A.M.E. Compensatory Growth, Feed Utilization, Whole-Body Composition, and Hematological Changes in Starved Juvenile Nile Tilapia, Oreochromis niloticus (L.) J. Appl. Aquac. 2006;18:17–36.



E-ISSN: 2229-7677 • Website: www.ijsat.org • Email: editor@ijsat.org

- 162. Mohanty M., Adhikari S., Mohanty P., Sarangi N. Effect of Waterborne Zinc on Survival, Growth, and Feed Intake of Indian Major Carp, Cirrhinus mrigala (Hamilton) Water. Air. Soil Pollut. 2009;201:3–7.
- 163. Palaniappan P.R., Nishanth T., Renju V.B. Bioconcentration of Zinc and Its Effect on the Biochemical Constituents of the Gill Tissues of Labeo rohita: An FT-IR Study. Infrared Phys. Technol. 2010;53:103–111.
- 164. Angon PB, Islam MS, Das A, Anjum N, Poudel A, Suchi SA. Sources, effects and present perspectives of heavy metals contamination: Soil, plants and human food chain. Heliyon. 2024 Apr 15;10(7).
- 165. Mishra S, Singh G, Gupta A, Tiwari RK. Heavy metal/metalloid contamination: their sources in environment and accumulation in food chain. InHeavy metal toxicity: environmental concerns, remediation and opportunities 2023 Sep 23 (pp. 19-47). Singapore: Springer Nature Singapore.
- 166. Tolkou AK, Toubanaki DK, Kyzas GZ. Detection of arsenic, chromium, cadmium, lead, and mercury in fish: Effects on the sustainable and healthy development of aquatic life and human consumers. Sustainability. 2023 Nov 23;15(23):16242.
- 167. Abbas K, Alam M, Kamal S. Heavy metals contamination in water bodies and its impact on fish health and fish nutritional value: A. International Journal of Fauna and Biological Studies. 2021;8:43-9.
- 168. Pandey G, Madhuri S. Heavy metals causing toxicity in animals and fishes. Research Journal of Animal, Veterinary and Fishery Sciences. 2014 Feb;2(2):17-23.
- 169. Isangedighi IA, David GS. Heavy metals contamination in fish: effects on human health. Journal of Aquatic Science and Marine Biology. 2019 Oct 31;2(4):7-12.
- 170. Cannon JR, Greenamyre JT. The role of environmental exposures in neurodegeneration and neurodegenerative diseases. Toxicological Sciences. 2011 Dec 1;124(2):225-50.
- 171. Campbell A. Inflammation, neurodegenerative diseases, and environmental exposures. Annals of the new york academy of sciences. 2004 Dec;1035(1):117-32.
- 172. Mohammadi MJ, Zarea K, Hatamzadeh N, Salahshouri A, Sharhani A. Toxic air pollutants and their effect on multiple sclerosis: a review study. Frontiers in Public Health. 2022 Jul 6;10:898043.
- 173. Roach K, Roberts J. A comprehensive summary of disease variants implicated in metal allergy. Journal of Toxicology and Environmental Health, Part B. 2022 Aug 18;25(6):279-341.
- 174. Parida L, Patel TN. Systemic impact of heavy metals and their role in cancer development: a review. Environmental Monitoring and Assessment. 2023 Jun;195(6):766.
- 175. Jomova K, Alomar SY, Nepovimova E, Kuca K, Valko M. Heavy metals: toxicity and human health effects. Archives of toxicology. 2025 Jan;99(1):153-209.
- 176. Masindi V, Muedi KL. Environmental contamination by heavy metals. Heavy metals. 2018 Jun 27;10(4):115-33.
- 177. Mishra S, Singh G, Gupta A, Tiwari RK. Heavy metal/metalloid contamination: their sources in environment and accumulation in food chain. InHeavy metal toxicity: environmental concerns, remediation and opportunities 2023 Sep 23 (pp. 19-47). Singapore: Springer Nature Singapore.
- 178. Guiné V., Spadini L., Sarret G., Muris M., Delolme C., Gaudet J.P., Martins J.M.F. Zinc Sorption to Three Gram-Negative Bacteria: Combined Titration, Modeling, and EXAFS Study. Environ. Sci. Technol. 2006;40:1806–1813.
- 179. Comte S., Guibaud G., Baudu M. Biosorption Properties of Extracellular Polymeric Substances (EPS) towards Cd, Cu and Pb for Different PH Values. J. Hazard. Mater. 2008;151:185–193.



- 180. Fang L., Wei X., Cai P., Huang Q., Chen H., Liang W., Rong X. Role of Extracellular Polymeric Substances in Cu(II) Adsorption on Bacillus subtilis and Pseudomonas putida. Bioresour. Technol. 2011;102:1137–1141.
- 181. Tálos K., Páger C., Tonk S., Majdik C., Kocsis B., Kilár F., Pernyeszi T. Cadmium Biosorption on Native Saccharomyces cerevisiae Cells in Aqueous Suspension. Acta Univ. Sapientiae Agric. Environ. 2009;1:20–30.
- 182. Tigini V., Prigione V., Giansanti P., Mangiavillano A., Pannocchia A., Varese G.C. Fungal Biosorption, an Innovative Treatment for the Decolourisation and Detoxification of Textile Effluents. Water. 2010;2:550–565.
- 183. Pinedo-Rivilla C., Aleu J., Collado I. Pollutants Biodegradation by Fungi. Curr. Org. Chem. 2009;13:1194–1214.
- 184. Gómez Jiménez-T R., Moliterni E., Rodríguez L., Fernández F.J., Villaseñor J. Feasibility of Mixed Enzymatic Complexes to Enhanced Soil Bioremediation Processes. Procedia Environ. Sci. 2011;9:54–59.
- 185. Penny C., Vuilleumier S., Bringel F. Microbial Degradation of Tetrachloromethane: Mechanisms and Perspectives for Bioremediation. FEMS Microbiol. Ecol. 2010;74:257–275.
- 186. Shahjahan M., Islam S.M.M., Bablee A.L., Siddik M.A.B., Fotedar R. Sumithion Usage in Aquaculture: Benefit or Forfeit? Rev. Aquac. 2021;13:2092–2111.
- 187. Sone Y., Mochizuki Y., Koizawa K., Nakamura R., Pan-Hou H., Itoh T., Kiyono M. Mercurial-Resistance Determinants in Pseudomonas Strain K-62 Plasmid PMR68. AMB Express. 2013;3:1–7.
- 188. Vutukuru S.S., Arun Prabhath N., Raghavender M., Yerramilli A. Effect of Arsenic and Chromium on the Serum Amino-Transferases Activity in Indian Major Carp, Labeo rohita. Int. J. Environ. Res. Public Health. 2007;4:224–227.
- 189. Pande V., Pandey S.C., Sati D., Bhatt P., Samant M. Microbial Interventions in Bioremediation of Heavy Metal Contaminants in Agroecosystem. Front. Microbiol. 2022;13:824084.
- 190. de Lorenzo V. Recombinant Bacteria for Environmental Release: What Went Wrong and What We Have Learnt from It. Clin. Microbiol. Infect. 2009;15:63–65.
- 191. Bhatt P., Sethi K., Gangola S., Bhandari G., Verma A., Adnan M., Singh Y., Chaube S. Modeling and Simulation of Atrazine Biodegradation in Bacteria and Its Effect in Other Living Systems. J. Biomol. Struct. Dyn. 2020;40:3285–3295.
- 192. Subashchandrabose S.R., Venkateswarlu K., Krishnan K., Naidu R., Lockington R., Megharaj M. Rhodococcus wratislaviensis Strain 9: An Efficient p-Nitrophenol Degrader with a Great Potential for Bioremediation. J. Hazard. Mater. 2018;347:176–183.
- 193. Barkay T., Miller S.M., Summers A.O. Bacterial Mercury Resistance from Atoms to Ecosystems. FEMS Microbiol. Rev. 2003;27:355–384.
- 194. Rojas L.A., Yáñez C., González M., Lobos S., Smalla K., Seeger M. Characterization of the Metabolically Modified Heavy Metal-Resistant Cupriavidus metallidurans Strain MSR33 Generated for Mercury Bioremediation. PloS ONE. 2011;6:e17555.
- 195. Yu L., Zhai Q., Zhu J., Zhang C., Li T., Liu X., Zhao J., Zhang H., Tian F., Chen W. Dietary Lactobacillus plantarum Supplementation Enhances Growth Performance and Alleviates Aluminum Toxicity in Tilapia. Ecotoxicol. Environ. Saf. 2017;143:307–314.
- 196. Zhai Q., Yu L., Li T., Zhu J., Zhang C., Zhao J., Zhang H., Chen W. Effect of Dietary Probiotic Supplementation on Intestinal Microbiota and Physiological Conditions of Nile Tilapia (Oreochromis



E-ISSN: 2229-7677 • Website: www.ijsat.org • Email: editor@ijsat.org

niloticus) under Waterborne Cadmium Exposure. Antonie van Leeuwenhoek, Int. J. Gen. Mol. Microbiol. 2016;110:501–513.

197. Ramadan A., Shalaby A., El Gammal M., El Aganif E.-S.M.E.A., Ebrahim M. Beneficial Using of EDTA to Reduce Cadmium Toxicity and to Improve the Physiological and Biochemical Profiles of Catfish (Clarias gariepinus) Egypt. J. Aquac. 2011;1:55–69.